



Product Information
Version 1.0

ZEISS LSM 800 with Airyscan Your Compact Confocal Power Pack



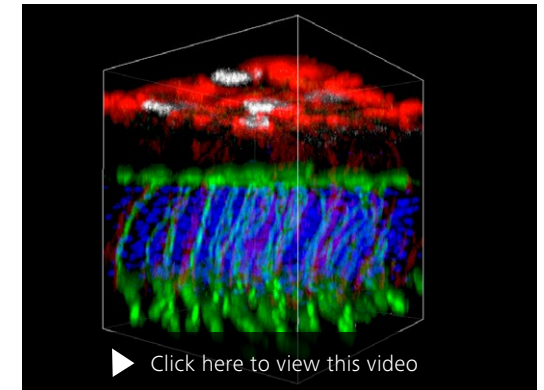
We make it visible.

Your Compact Confocal Power Pack

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Confocal imaging demands the very best imaging quality. With LSM 800 you are choosing a flexible and compact confocal laser scanning microscope, complete with highly sensitive GaAsP detector technology and fast linear scanning. Add Airyscan, the revolutionary detection concept from ZEISS, and you will gain 1.7× higher resolution in all three dimensions – resulting in a 5× smaller confocal volume. And you will be pushing sensitivity beyond the limits of all conventional confocals.

LSM 800 is your entry into the world of high-end confocal imaging. Simply decide which options your system needs today, then upgrade in the future as your needs grow.



Mouse retina. Mueller cells stained for RFP (red), Amacrine cells stained for Chat (green), Cone photo receptors stained for mCar (white) and DNA stained with Hoechst (blue); Sample: courtesy of B. Roska, Friedrich Miescher Institute for Biomedical Research, Basel, Switzerland

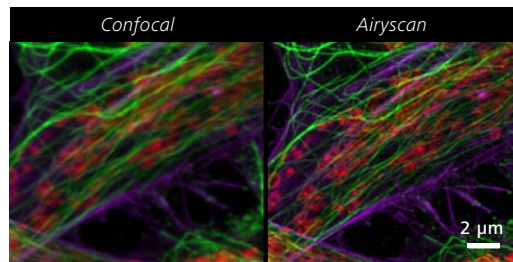
See for yourself how LSM 800 with Airyscan will increase your productivity. Book a hands-on demonstration in one of our ZEISS Microscopy Labs now. >> www.zeiss.com/lsm800

Simpler. More Intelligent. More Integrated.

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Perfectly Tailored to Your Needs

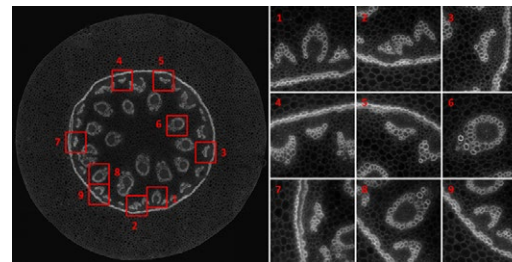
With up to three highly sensitive GaAsP detectors and fast linear scanning, LSM 800 brings you higher productivity and throughput, greater flexibility in live cell imaging and uncompromised image quality. Use this confocal power pack for precise quantitative measurements. Then take advantage of Airyscan, the revolutionary detection concept, for 1.7 times higher resolution and higher sensitivity than any classic detection method can deliver.



Comparison between confocal and Airyscan image. HeLa cells, red: mitochondria membran, green: microtubuli, magenta: actin fibers. Sample: courtesy of A. Seitz, Biolmaging and Optics Core Facility, EPFL, Lausanne, Switzerland.

Use Open Interfaces to Extend Your System

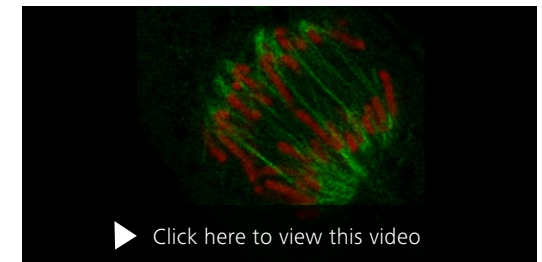
Give your lab or multi-user facility the full benefits of integrated incubation solutions and state-of-the-art Axiocams. LSM 800 uses intuitive ZEN imaging software for complex automated imaging routines with Experiment Designer. Yet it's just as easy to exchange data with third party software and define your own application world using the powerful Open Application Development (OAD). ZEISS Shuttle & Find for correlative microscopy connects LSM 800 with your ZEISS electron microscope.



Open Application Development (OAD) is the open and well-documented Python interface for ZEN imaging software. The example shows rare event detection whereby the scan is analyzed and interesting regions re-scanned at high resolution.

Your Compact System for High-end Confocal Imaging

LSM 800 makes excellent economic sense: an affordable system with an attractive price / performance ratio. It's robust and easy to use, with a small footprint and minimal setup requirements – combined with minimal maintenance, minimal training, self-calibration and low energy consumption. That adds up to a predictable cost of ownership over its entire lifetime.



Culture of living LLC-PK1 (Pig Kidney Epithelial) cells, green: tubulin-GFP, red: H2B-mCherry.

Your Insight into the Technology Behind It

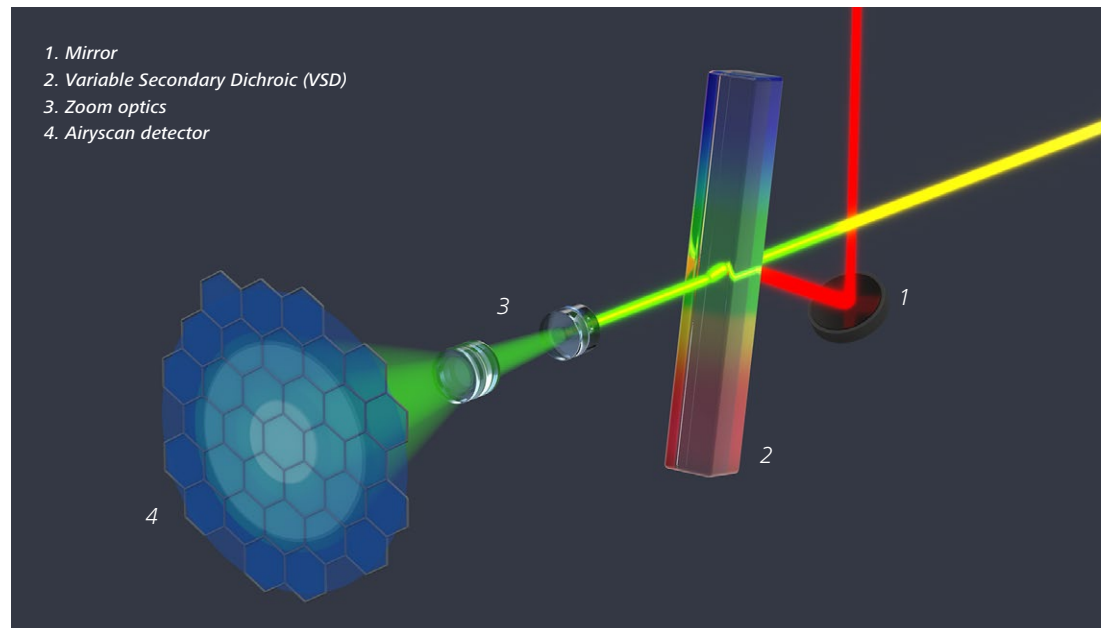
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Revolutionize Your Confocal Imaging with ZEISS Airyscan

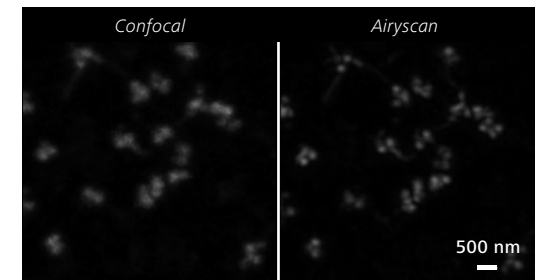
Airyscan is an array detector that draws on the fact that a fluorescence microscope will image a point-like source as an extended Airy disk. When you close the pinhole in a standard confocal microscope to reject out-of-focus light, you get a sharper image, but it's also dimmer since a great deal of light is then lost. The smaller the pinhole, the higher the resolution, but – equally – the bigger the loss in light.

Airyscan solves this conundrum between resolution and light efficiency by imaging the complete Airy disk onto a concentrically arranged hexagonal detector array. It consists of 32 single detector elements, all of which act like sub-Airy unit pin-holes. The confocal pinhole itself remains open and doesn't block light – thus all photons of the whole Airy disk are collected. The signals from all detector elements are then reassigned to their

correct position, producing an image with increased signal-to-noise ratio and resolution. Unlike other superresolution techniques, Airyscan capitalizes on the scanning and optical sectioning capabilities of a confocal. Thus Airyscan even works with thicker samples such as tissue sections or whole animal mounts that need a higher penetration depth.



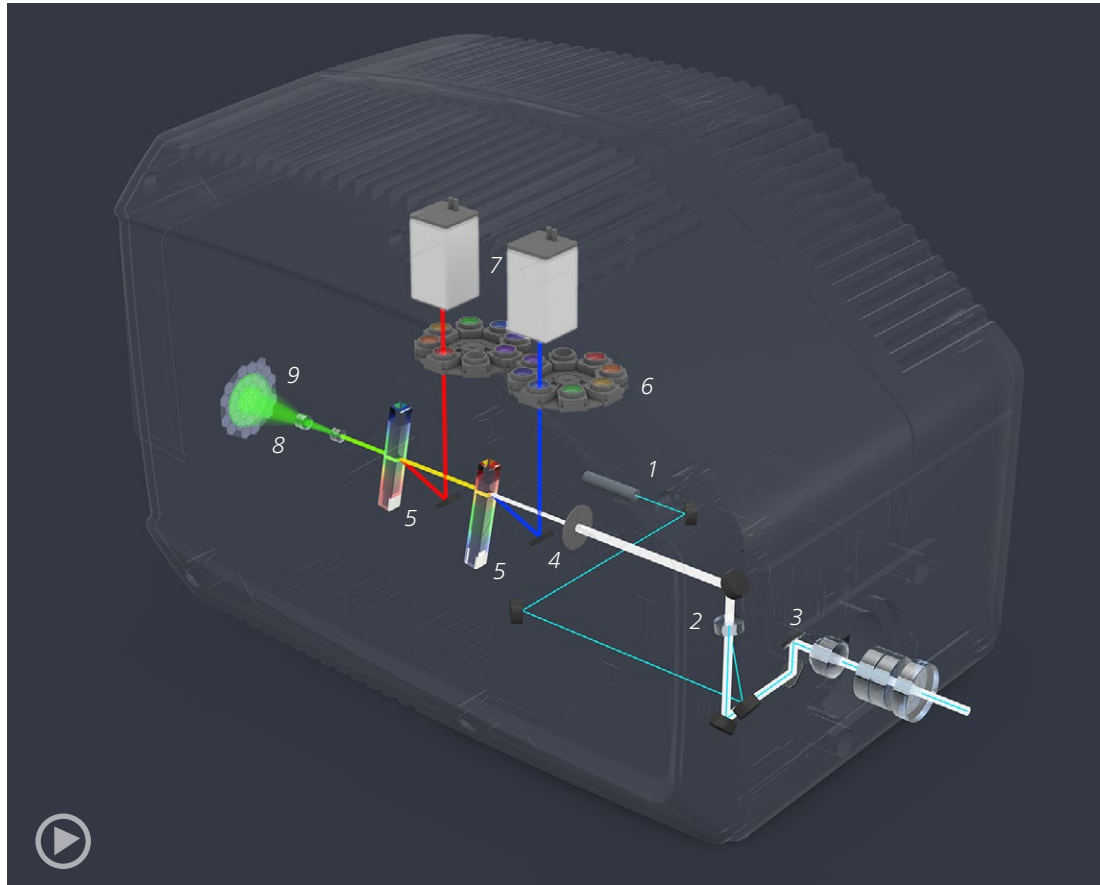
Schematic beam path of Airyscan.



Isolated centrioles of *Chlamydia*, fixed with Methanol; Tubulin staining with Alexa 488. Sample: courtesy of P. Guichard, EPFL, Lausanne, Switzerland.

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Schematic beam path of LSM 800

- | | |
|--------------------------------------|-----------------------|
| 1. Excitation laser lines | 6. Emission filters |
| 2. Main beam splitter (MBS) | 7. Confocal detectors |
| 3. Galvo scanning mirrors | 8. Zoom optics |
| 4. Pinhole | 9. Airyscan detector |
| 5. Variable Secondary Dichroic (VSD) | |

A Streamlined Light Path with Surprising Flexibility

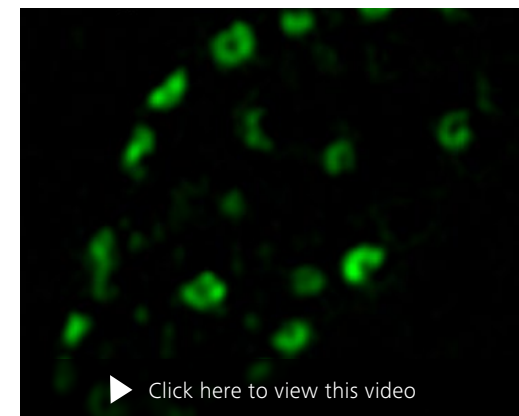
The compact light path with a minimum of optical elements is designed for highest efficiency. Fluorescence emission light travels through the main dichroic beam splitter with its outstanding laser suppression to deliver supreme contrast. Up to two patented variable beam splitter dichroics (VSDs) divert the spectral part of the light. You can freely define up to three detectors (multialkali, GaAsP or Airyscan).

Your Insight into the Technology Behind It

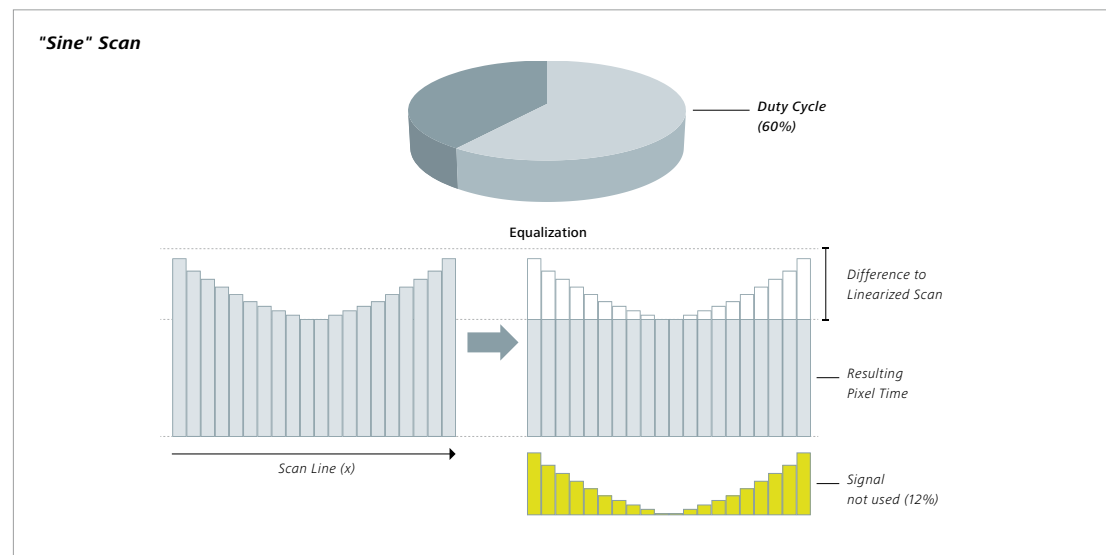
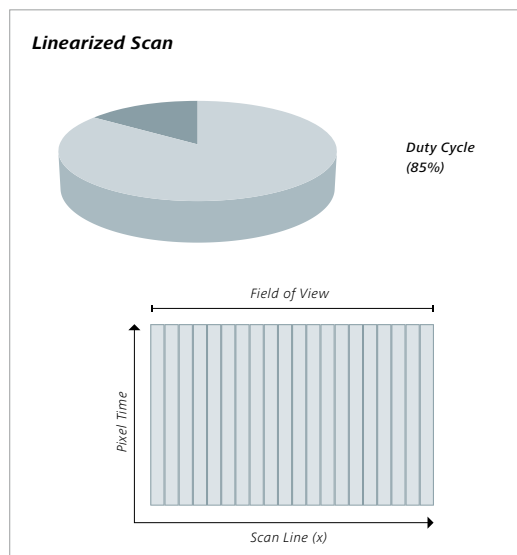
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Fast and Linear Scanning – Your Powerful Combination

LSM 800 gives you the benefit of increased scanning speeds so you can resolve those fast movements of labeled proteins that demand equally fast scanning. At an image size of 512×512 pixels you will be capturing events with up to 8 frames per second. Your LSM 800 is constantly monitoring and calibrating the scanner position so you can count on a stable and even field of view with constant pixel times across the whole observation area. This patented linear scanning regime gives you a constant signal-to-noise level and uniform exposure by the illuminating laser throughout the scanned area, including your manipulated regions of interest. With LSM 800 you will be using more than 80% of the scanning time for data acquisition. Signal-to-noise improves by about 29% compared to sine scanning systems. Your experiments will always deliver quantitative data. Likewise, you can adapt the scan field at any time by panning or cropping it, and rotating it freely to best suit the geometry of your sample.



Drosophila melanogaster neuromuscular junction stained for Bruchpilot (BRP). Comparison between confocal GaAsP and Airyscan detection. Sample: courtesy of J. Pielage, Friedrich Miescher Institute for Biomedical Research, Basel, Switzerland.



Your Insight into the Technology Behind It

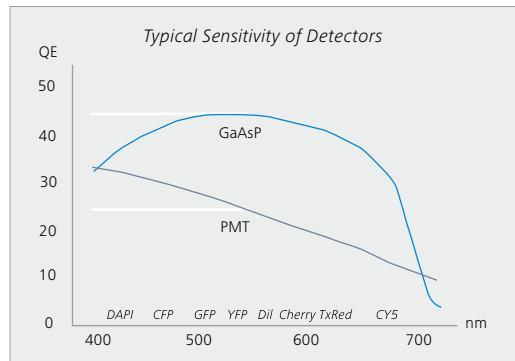
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GaAsP Detectors – Your Choice for Highest Sensitivity

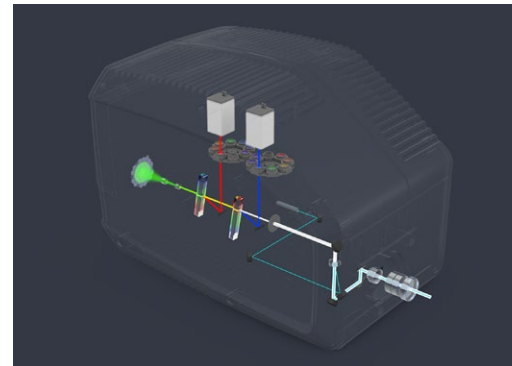
GaAsP PMTs – that is, gallium arsenide phosphide photomultiplier tubes – display high light collection efficiencies over a broad spectral range. Their low dark noise levels also render them the ideal tool for detecting faint signals. Enjoy outstanding image quality based on a superb signal-to-noise ratio (SNR). You might use this gain in SNR to increase productivity by achieving faster scan speeds while preserving excellent image quality. Or take advantage of the low laser powers needed in live cell imaging applications to avoid photobleaching and phototoxicity as much as possible. Or simply detect faint signals in low expressing cells. All that, and you can do it with up to three spectral channels simultaneously.

Benefit from up to Three Confocal Detectors

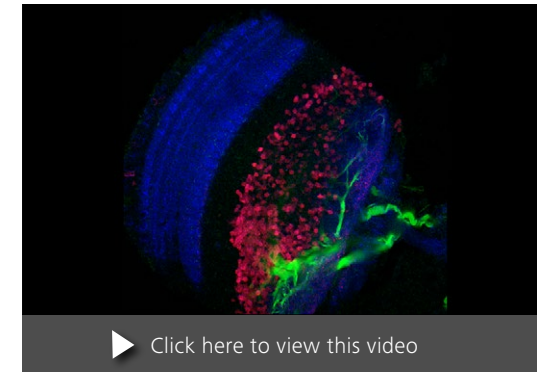
Investigations into localization and interaction of proteins often require multiple fluorescent labels with overlapping emission spectra. Now you can image up to four dyes, crosstalk free by multitracking. Or even more by performing a Lambda scan with spectral unmixing.



Typical spectral quantum efficiency (QE) of PMT and GaAsP detectors.



Schematic beam path of LSM 800.



[Click here to view this video](#)

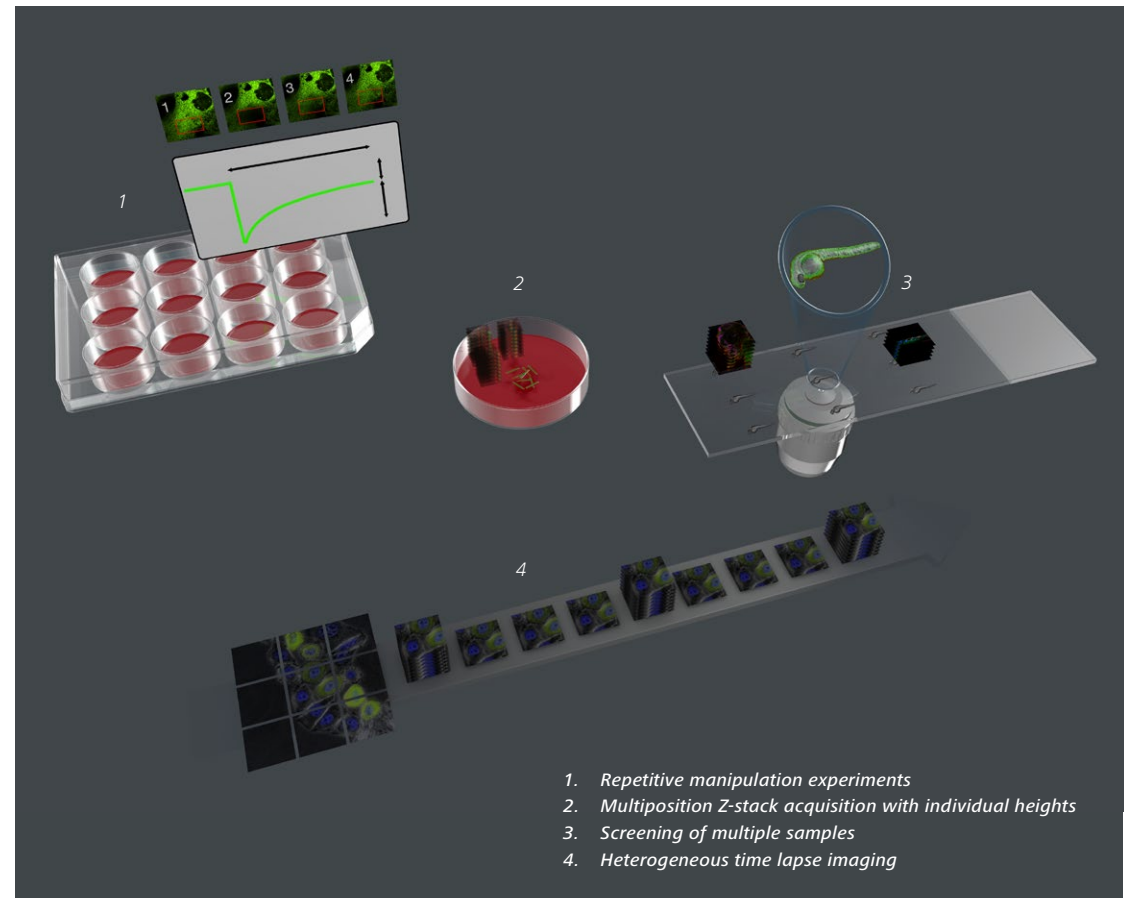
Drosophila brain; triple antibody staining: Alexa 488, Alexa 568 and Alexa 633; Sample: courtesy of D. Reiff, Institute of Biology, Albert-Ludwigs-University Freiburg, Germany.

Expand Your Possibilities

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Experiment Designer: Your Smart Automation Module for Enhanced Productivity

Use Experiment Designer to automate complex acquisition strategies. Exploit and combine different imaging modalities. Execute repetitive imaging of a large number of samples. In this way, you will get results that are statistically validated.

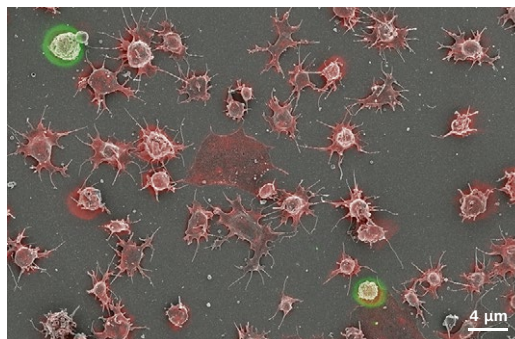
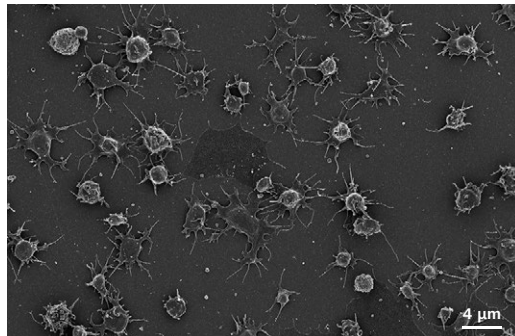
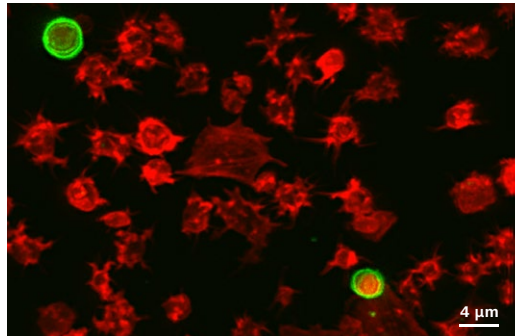


With the ZEN software module Experiment Designer you can set up complex imaging routines consisting of freely defined and repeatable experiment blocks with multi-position tile scans of multichannel Z-stacks.

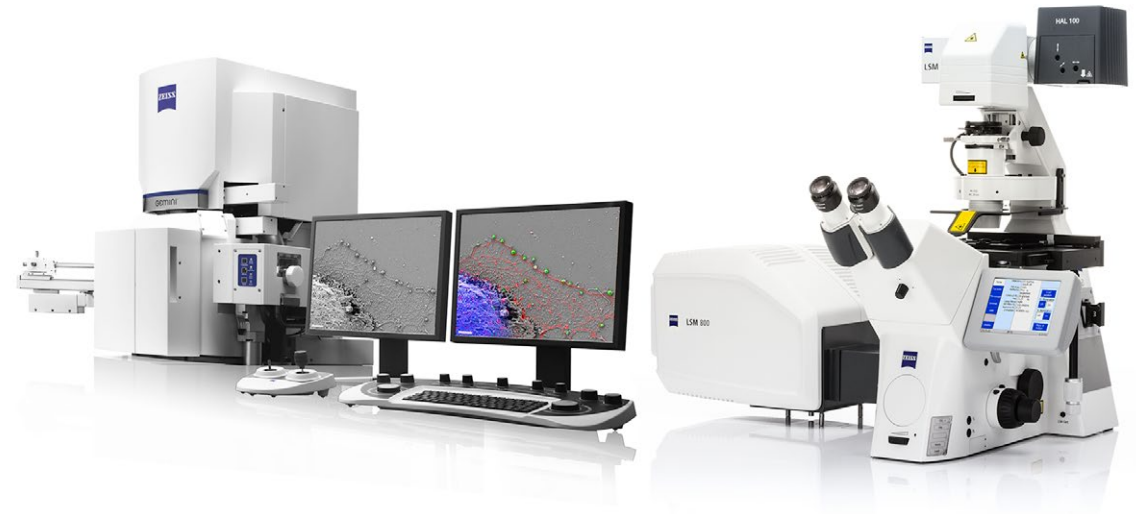
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Correlative Microscopy



Platelets stained for cellular platelet protein (green) and actin (red). Upper image: LSM fluorescence image; center image: SEM image; bottom image: overlay. Courtesy of D. Woulfe and J. Caplan, University of Delaware, Newark, USA.



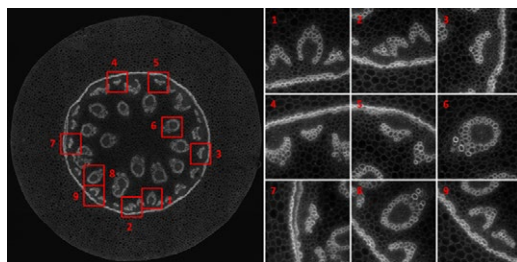
To map the distribution of fluorescently labeled proteins to subcellular structures with the highest precision, the Shuttle & Find module is your technology of choice. A wizard-guided easy-to-use workflow between light and scanning electron microscope delivers reliable relocation of defined regions of interest. Images from both microscopical methods can be overlaid to one correlative image revealing functional information within an ultrastructural context.

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OAD is Your Interface to ZEN Imaging Software

- Use Python scripts to customize and automate your workflows.
- Integrate external image analysis applications into your workflow.
- Exchange image data with external programs like ImageJ, Fiji, MATLAB, KNIME or Python.
- Use feedback for smart experiments.
- Get more reliable data in less time. It's your choice.



Rare event detection. A Convallaria sample was scanned and the image analyzed for features. Areas with hits were re-scanned at higher magnification.



OAD enables the analysis of data acquired with ZEN Imaging Software by other programs like ImageJ. Transfer your results back to ZEN for further analysis and display.

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As your needs grow, LSM 800 grows with you, forming the basis for a number of enhancements. Like every system from ZEISS, LSM 800 comes with open interfaces and a modular architecture to guarantee the seamless interaction of all components, now and in the future.



Combine Axio Observer with incubation to get the best tool for long-term live cell imaging with stable temperature conditions.



Add the newest choice of cameras from the AxioCam series to LSM 800 for widefield imaging experiments – also in combination with LSM imaging.



Shuttle & Find is your gateway to correlative light and electron imaging (CLEM). Combine the specificity of functional fluorescence imaging with ultrastructural information.



Z piezo stage and a leveling insert guarantee the precision needed for superresolution applications using Airyscan.



Definite Focus stabilizes the focal position of your sample compensating z-drift. You can now perform long-term experiments that can last for multiple days.

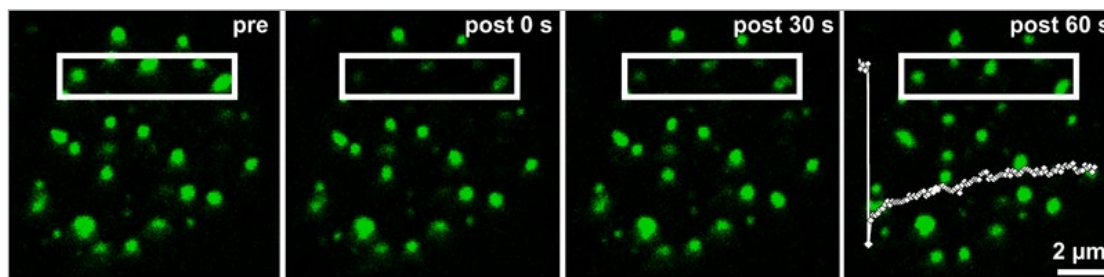


The electronically switchable illumination and detection module (ESID) combines transmitted light illumination and detection in one component. No mechanical parts need to be moved when switching between modes.

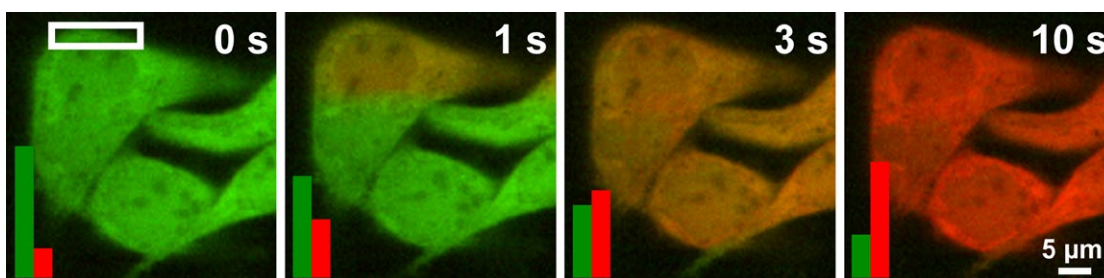
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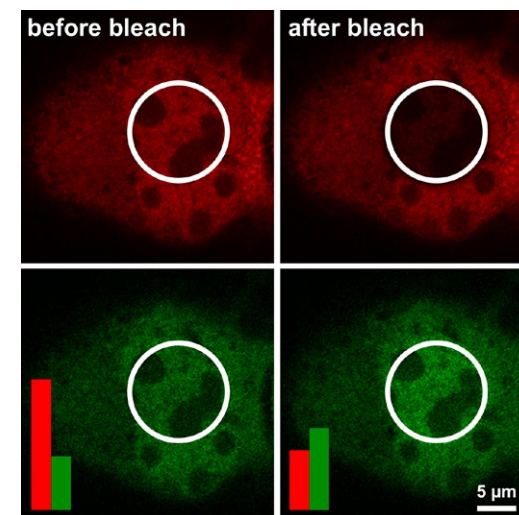
Use the outstanding sensitivity of LSM 800 for the study of protein dynamics in living cells.



Use FRAP to study protein dynamics. The example shows EGFP-CENPI in HepG2 cells before bleach ("pre"), and at the indicated time points after the bleach ("post"). The recovery curve (superimposed in the last image and showing the recovery from bleach at 0 s to 60 s, intensities in AU) can be used to calculate the diffusion coefficient of the molecule.



Photoactivation is your method of choice to study the fate of proteins. The example shows Kaede expressed in HepG2 cells before photoactivation (0 s) and at different time points (1 s, 3 s and 10 s) after repeated photoactivation (every 0.1 s) with 405 nm at the indicated regions (white box). Kaede diffuses freely between the nucleus and the cytoplasm. The relative intensities in AU of the non-converted form (green bars) and converted form (red bars) are shown in each image.



FRET is your tool for investigation of protein interaction. The example shows two interacting proteins (donor false colored in green, acceptor false colored in red) in HepG2 cells ("before bleach"). By acceptor-photobleaching (within the indicated white circle) acceptor intensity will decrease, while donor intensity will increase ("after bleach") as indicated by the green (donor) and red (acceptor) bars. The increase in donor intensity can be used to calculate FRET efficiencies.

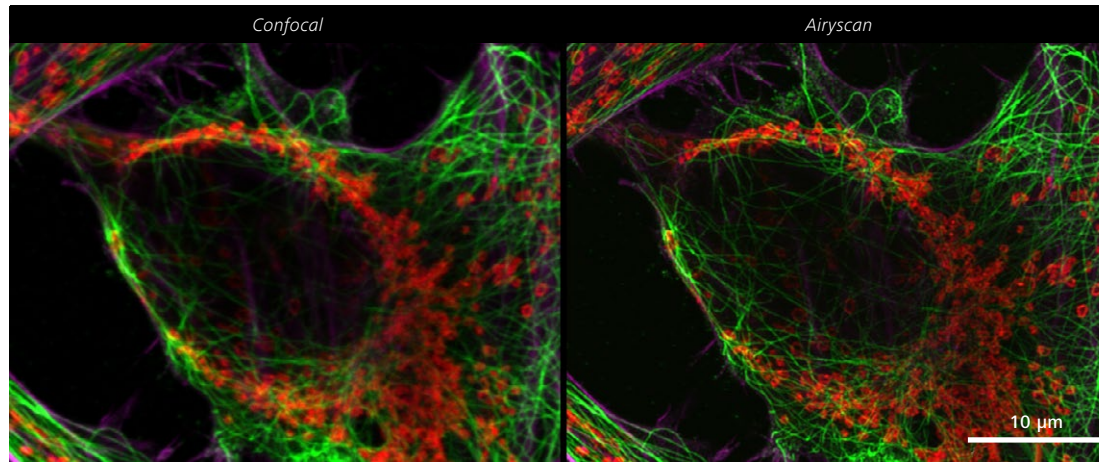
Tailored Precisely to Your Applications

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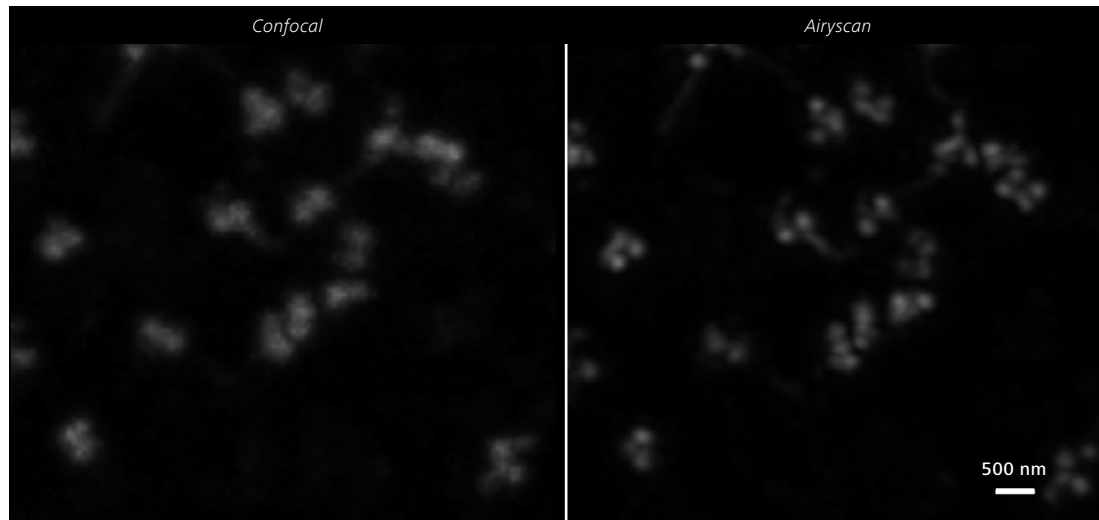
Typical Applications, Typical Samples	Task	ZEISS LSM 800 Offers
Antibody stained tissue slices	Document morphological relations of structures with a resolution of 140 nm (xy) / 400 nm (z) at 488 nm excitation	Airyscan with GaAsP detector for superresolution imaging
Live cell culture	Study the motility of vesicles and organelles	Up to 8 frames per second time lapse imaging
	Document the kinetics of endo- and exocytosis	Mixed mode imaging with LSM and superresolution based on photoactivated localization
	Screen and document cells expressing the desired fluorescent label in response to pharmacological treatment	Widefield imaging using AxioCam
Live cell culture with two labels	Study the motility of subcellular structures	Airyscan with GaAsP detector to image with time lapse imaging in 2D or 3D at 1.6 frames per second
	Explore the interaction of two proteins exploiting the Förster Resonance Energy Transfer effect	ZEN (black edition) FRET analysis
Live cells with multiple labels	Image over a long time in an automated way	Experiment Designer software tool combined with three parallel spectral channels
Live or fixed cells with multiple labels and overlapping emission signals	Examine the interplay of multiple proteins	Parallel acquisition of all signals with three spectral channels and linear unmixing
Cellular structures with weak labels	Image subcellular structures at physiological expression levels	Airyscan with GaAsP detector or LSM 800 with GaAsP detector
Study molecular dynamics	Photomanipulation	ZEN (black edition) FRAP analysis
Plant roots	Follow the changes of subcellular structures over time with high resolution	Airyscan with GaAsP detector for superresolution imaging beyond 40 µm deep into tissue with up to 1.6 full frames per second (512 × 512 pixels)
Model organisms, e.g. Zebrafish, <i>Drosophila</i> or <i>C. elegans</i>	See fine details of the organization and dynamics of endogenously expressed FP proteins	Airyscan with GaAsP detector for superresolution imaging beyond 40 µm deep into tissue

ZEISS LSM 800 at Work

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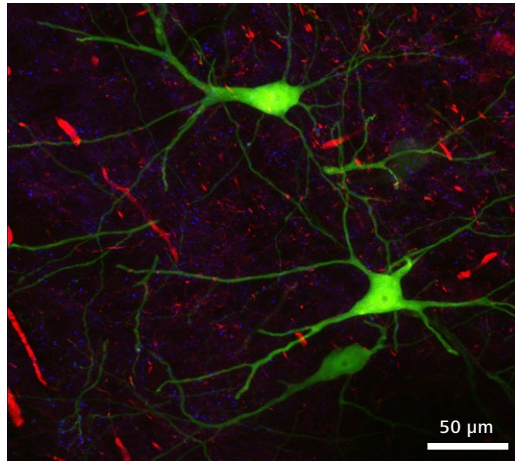
Comparison between confocal and Airyscan image. HeLa cells, red: mitochondria membrane, green: microtubuli, magenta: actin fibers. Sample: courtesy of A. Seitz, Bioluminescence and Optics Core Facility, EPFL, Lausanne, Switzerland.



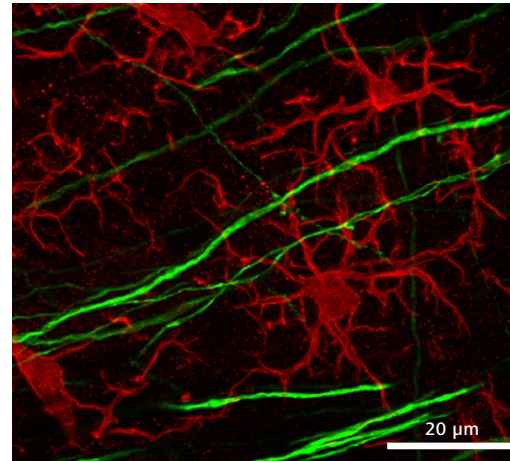
Isolated centrioles of Chlamydia; fixed with Methanol; Tubulin staining with Alexa 488. Sample: courtesy of P. Guichard, EPFL, Lausanne, Switzerland.

ZEISS LSM 800 at Work

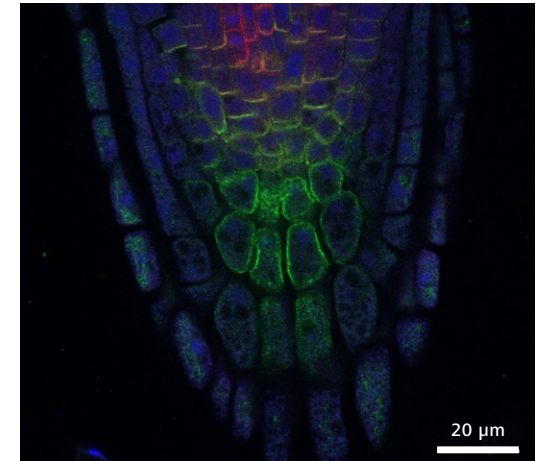
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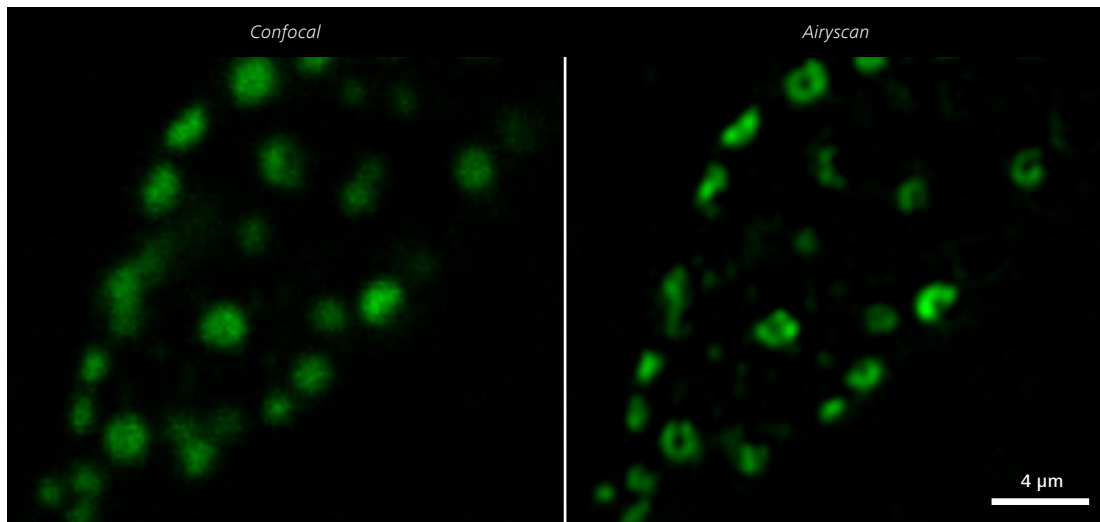
Mouse brain slice, EGFP-Thy1 (green): nerve cells (subset), Calretinin-Cy3 (red): Calretinin-expressing neurons, GAD65-Cy5 (blue): GABAergic synapses; Sample: courtesy of P. Janz, Neuropathology, University Freiburg, Germany.



Mouse brain slice, EGFP-Thy1: nerve cells, Iba1-Cy3: microglia cells; Sample: courtesy of P. Janz, Neuropathology, University Freiburg, Germany.



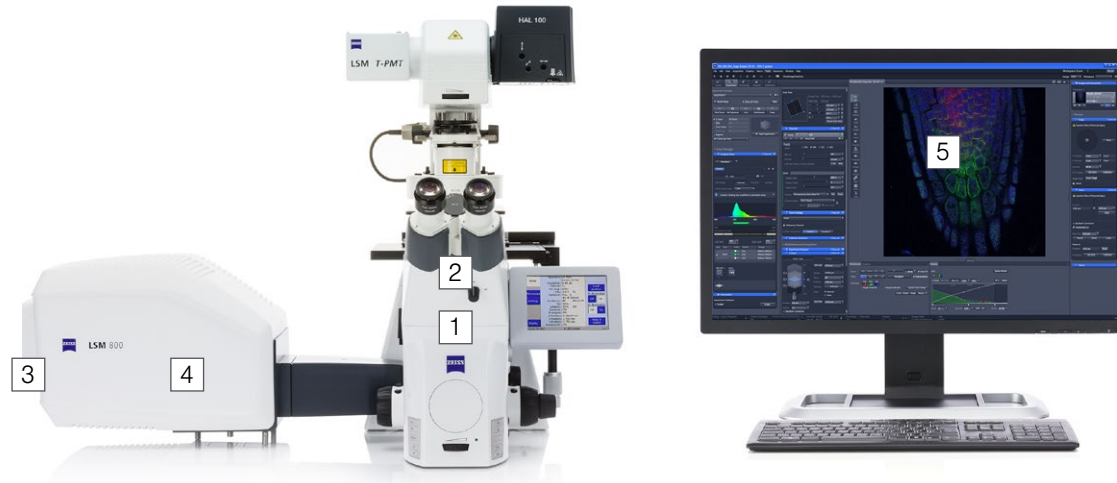
Arabidopsis thaliana root, PIN1 (red), PIN4 (green), DAPI (blue); Sample: courtesy of T. Pasternak, Institute of Biology, Albert Ludwigs University Freiburg, Germany.



Drosophila melanogaster neuromuscular junction stained for Bruchpilot (BRP). Comparison between confocal GaAsP (left) and Airyscan (right) detection. Sample: courtesy of J. Pielage, Friedrich Miescher Institute for Biomedical Research, Basel, Switzerland.

ZEISS LSM 800: Your Flexible Choice of Components

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1 Microscope

- Inverted stand: Axio Observer
- Upright stand: Axio Imager
- Camera port
- Manual or motorized stages
- Incubation solutions
- Fast Z piezo inserts (for inverted stands)
- Definite Focus

2 Objectives

- C-APOCHROMAT
- Plan-APOCHROMAT
- LD Plan-APOCHROMAT
- EC Plan-NEOFLUAR

3 Illumination

- Diode lasers: 405, 488, 561 and 640 nm

4 Detection

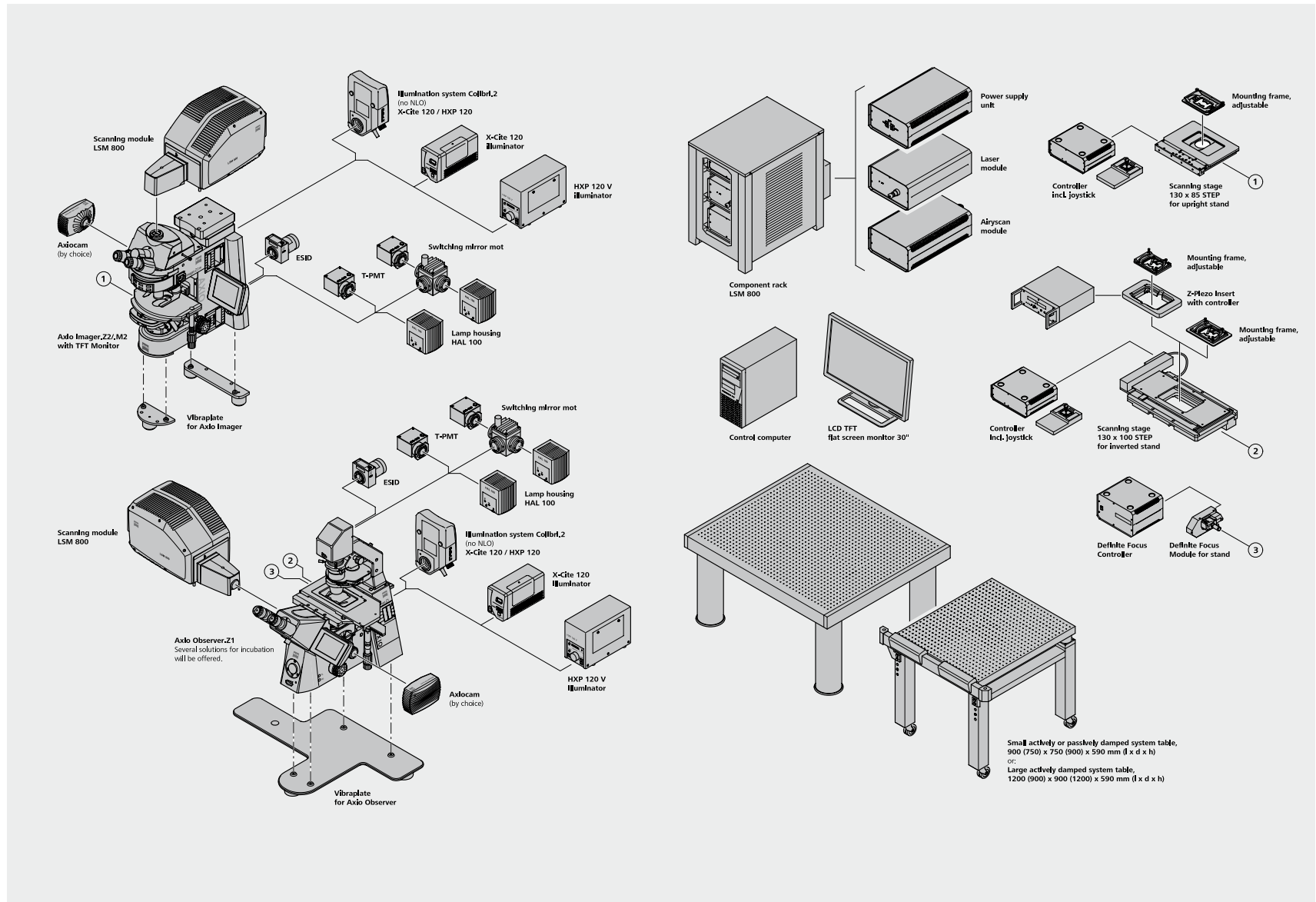
- 2 channel Gallium Arsenide Phosphid (GaAsP) PMT or 2 channel multialkali (MA) PMT
- 1 additional GaAsP PMT, multialkali PMT or Airyscan detector for 40x or 63x objective
- Electronically switchable illumination and detection module (ESID) or transmitted light detector (T-PMT) with halogen lamp (HAL)

5 Software

- ZEN (blue edition), recommended modules:
Tiles & Positions, Experiment Designer,
3D VisArt

ZEISS LSM 800: System Overview

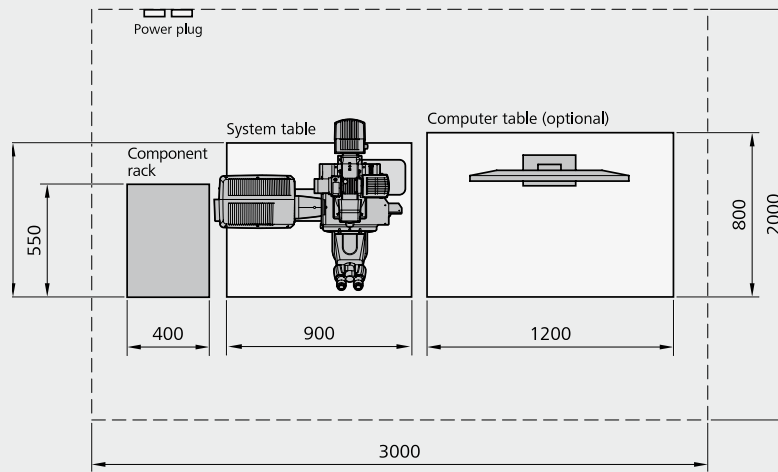
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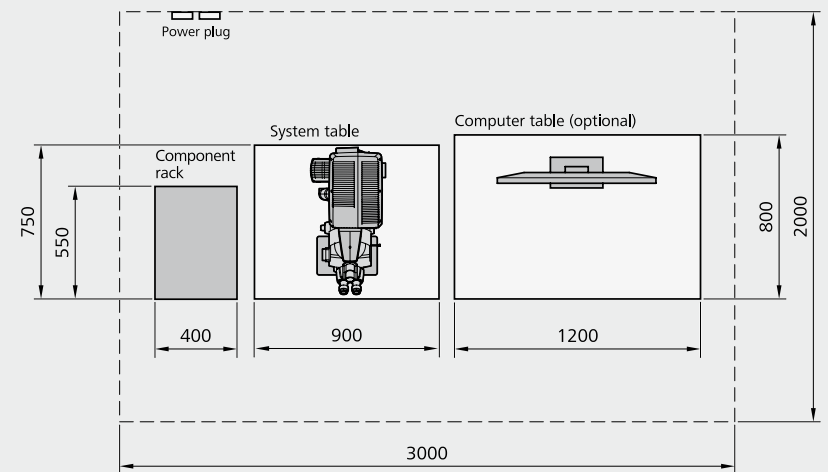
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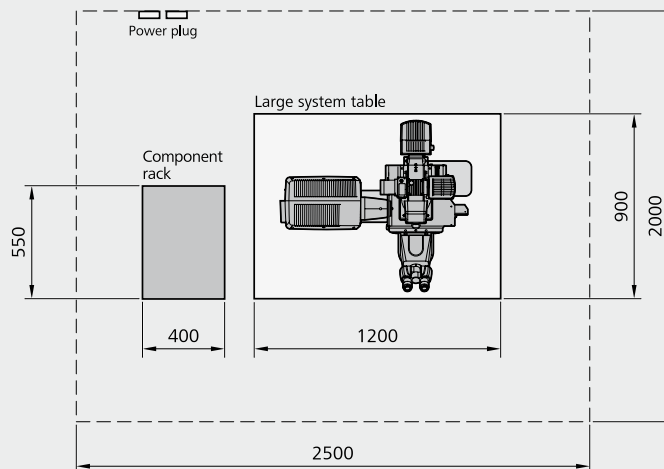
LSM 800 with Axio Observer on small system table



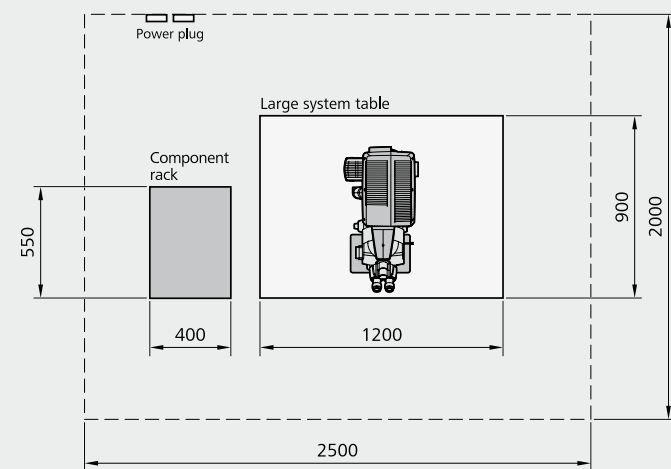
LSM 800 with Axio Imager on small system table



LSM 800 with Axio Observer on large system table



LSM 800 with Axio Imager on large system table



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Physical Dimensions	Length (cm)	Width (cm)	Height (cm)	Weight (kg)
Small actively and passively damped system table	75	90	59	170
Large actively damped system table	90	120	59	197
Vibraplate for Axio Imager (consists of three pedestals)	32	30	4.5	1.5
Vibraplate for Axio Observer	52.5	80	4.5	7
Scanning Module LSM 800	40	25.5	28	15
Axio Imager.Z2; Axio Imager.M2	56	39	70	20
Axio Observer.Z1	61	39	65	20
Component rack	55	40	60	35
Laser module (LM)	40	25	14.5	10
Airyscan (40x and 63x)	40	25	14.5	5
Power supply unit (PSU)	40	25	14.5	6
Fiber optic cable, VIS(ible)	300			
Cables	300			
Microscopes				
Stands	Upright: Axio Imager.Z2, Axio Imager.M2 Inverse: Axio Observer.Z1 with side port			
Z Drive	Smallest increment Axio Observer.Z1 and Axio Imager.M2: <25 nm; Axio Imager.Z2: <10 nm; Z-Piezo stage available; Definite Focus for Axio Observer.Z1			
XY Stage (optional)	Motorized XY scanning stage, for Mark & Find function (xy) as well as Tile Scan (Mosaic Scan) (Tiling not available for Airyscan detection); smallest increment of 1 µm (Axio Observer) or 0.2 µm (Axio Imager)			

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Scanning Module	
Scanner	Two independent, galvanometric scanning mirrors with ultrashort line and frame flyback
Scanning Resolution	4 × 1 to 6,000 × 6,000 pixels (Airyscan max. 4,000 × 4,000 pixels), also for multiple channels, continuously adjustable (for each axis)
Scanning Speed	Up to 8 images/sec (Airyscan up to 1.6 images/sec) with 512 × 512 pixels; up to 64 images/sec with 512 × 64 pixels
Scanning Zoom	0.5x to 40x; continuously adjustable
Scanning Rotation	Can be rotated freely (360 degrees), adjustable in increments of one degree, freely adjustable xy offset
Scanning Field	12.7 mm × 12.7 mm in the intermediate image plane, with full pupil illumination
Pinhole	Master pinhole with preset size and position; can be adjusted as desired for multitracking and short wavelengths (such as 405 nm); automatic alignment
Beam Path	One major beam splitter for four laser lines (405, 488, 561 and 640 nm) at 10 degree with excellent laser line suppression. The 640 nm laser line can be used for internal autofocusing. Depending on the system, either one or two patented Variable Secondary Dichroics (VSDs) can be used to flexibly divert the respective spectral range of light to chosen channels. Emission filters can be used to clean up the signal when imaging autofluorescent or highly scattering samples.
Detection Options	
Detectors	2 spectral detection channels, GaAsP (typical QE 45%) or multialkali (MA) PMT (typical QE 25%) 1 additional GaAsP PMT, multialkali PMT or Airyscan detector Airyscan with spatial detection (32 channels GaAsP) adapted for 40x or 63x objectives Transmitted light detector (ESID or T-PMT)
Spectral Detection	>8 sequential confocal fluorescence channels, GaAsP or MA based; up to three parallel confocal fluorescence channels, GaAsP or MA based
Data Depth	8-bit or 16-bit available
Real-Time Electronics	Microscope, laser, scanning module and additional accessory control; data acquisition and synchronization management through real-time electronics; oversampling read-out logic for best sensitivity; data transfer between real-time electronics and user PC via LVDS with the ability to evaluate the data online during image acquisition

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ZEN Imaging Software	
GUI Configuration	Workspace to conveniently configure all of the motorized functions of the scanning module, laser and microscope; save and restore application configurations (re-use)
Calibration Tools	Calibration objective and software tools to calibrate the system
Recording Modes, Smart Setup	Line, Frame, Tiles (not available for Airyscan), Z Stack, Lambda Stack, Time Series and all combinations (XYZ, lambda, t), online calculation and visualization of ratio images, average and summation (by line/image, adjustable), Step Scan (for higher image frame rates); quick setup of imaging conditions using Smart Setup by simply selecting the labeling dye
Crop Function	Easily select scanning areas (simultaneously select zoom, offset, rotation)
Real ROI Scan, Spline Scan	Scans of up to 99 designated ROIs (regions of interest) as desired and pixel-by-pixel laser blanking; scan along a freely defined line
ROI Bleaching	Localized bleaching in up to 99 bleach ROIs for applications such as uncaging; use of different speeds for bleaching and imaging, use of different laser lines for different ROIs
Multitracking	Rapidly change excitation lines when recording multiple fluorescences for the purpose of minimizing signal crosstalk and increasing dynamic range
Lambda Scan	Sequential acquisition of image stacks with spectral information for every pixel
Linear Unmixing	Acquisition of crosstalk-free, multiple fluorescence images using simultaneous excitation; offline unmixing; advanced unmixing logic with indication of reliability
Visualization	XY, orthogonal (XY, XZ, YZ), Cut (3D section); 2.5D for time series of line scans, projections (maximum intensity); animations; depth coding (inverse colors), brightness, gamma and contrast settings; color table selection and modification (LUT), character functions
Image Analysis and Operations	Co-localization and histogram analysis with individual parameters, profile measurement along user-defined lines, measurement of lengths, angles, areas, intensities and much more; operations: addition, subtraction, multiplication, division, ratio, shift, filters (low-pass, median, high-pass, etc., also user-definable)
Image Management	Features for managing images and the corresponding imaging parameters; multiprint feature
Optional Software	
3D VisArt	Rapid 3D and 4D reconstructions and animations (available modes: shadow projections, transparency projection, surface rendering)
Deconvolution	3D image restoration based on calculated point-spread functions (modes: nearest neighbor, maximum likelihood, constrained iterative)
Dynamics	Comprehensive evaluation software for online and offline calibration of ion concentrations
Open Application Development (OAD)	Python scripting interface for automation & customization; Experiment Feedback for Smart Experiments and open interface to third party software (e.g. ImageJ);
Experiment Designer	Definition of advanced automated imaging

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Lasers	
Laser Module URGB (pigtailed; 405, 488, 561, 640 nm)	<p>Single-mode polarization preserving fiber</p> <p>Direct modulation (500:1); for LSM 800 GaAsP system additional attenuation by a factor of 20 available to yield a dynamic range of around 10,000:1</p> <p>Diode laser (405 nm, 5 mW); laser class 3B</p> <p>Diode laser (488 nm, 10 mW); laser class 3B</p> <p>Diode (SHG) laser (561 nm, 10 mW); laser class 3B</p> <p>Diode laser (640 nm, 5 mW); laser class 3B</p>
Laser Module GB (pigtailed; 488, 561 nm)	<p>Single-mode polarization preserving fiber</p> <p>Direct modulation (500:1); for LSM 800 GaAsP system additional attenuation by a factor of approximately 20 available</p> <p>Diode laser (488 nm, 10 mW); laser class 3B</p> <p>Diode (SHG) laser (561 nm, 10 mW); laser class 3B</p>

Power Requirements

LSM 800 has a main power supply cord and country specific or plug NEMA L 14-30P (2/N/Ground 120/240V/30A) plug, and the matching mains socket outlet. The mains socket outlet must be equipped with a fuse having minimum tripping characteristic C according to IEC/EN 60898.

Line Voltage	100 V AC ... 125 V AC (+10%)	220 V AC ... 240 V AC (+10%)
Line Frequency	50 ... 60 Hz	50 ... 60 Hz
Max. Current	1 phase at 5 A	2 phases at 3 A
Power Plug	NEMA 5/15 (only USA)	Country specific connectors
Power Consumption	550 VA (continuous operation; maximum)	575 VA (continuous operation; maximum)
	260 VA (standby operation)	280 VA (standby operation)
	0.011 VA (off mode)	0.025 VA (off mode)
Heat Emission	500 W	500 W

EMC Test

according to DIN EN 61326-1 (07/2013)

1. Noise emission according to CISPR 11 / DIN EN 55011 (04/2011)
2. Noise immunity according to table 2 (industrial sector)

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Environmental Requirements

For operation, the system has to be placed in a closed room.

1. Operation, specified performance	T = 22° C ±3° C without interruption (24 h a day independently whether system is operated or switched off) It has to be ensured that the airflow of the air-conditioning is not directed at the system.
2. Operation, reduced performance	T = 15° C to 35° C, any conditions different from item 1. and 4.
3. Storage, less than 16 h	T = -20° C to 55° C
4. Temperature gradient	±0.5° C/h
5. Warm-up time	1 h for standard imaging; ≥2h for high-precision and/or long-term measurements
6. Relative humidity	<65% at 30° C
7. Operation altitude	max. 2,000 m
8. Loss of heat	500 W



LSM 800 meets the requirements according to IEC 60825-1:2007

Count on Service in the True Sense of the Word

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Because the ZEISS microscope system is one of your most important tools, we make sure it is always ready to perform. What's more, we'll see to it that you are employing all the options that get the best from your microscope. You can choose from a range of service products, each delivered by highly qualified ZEISS specialists who will support you long beyond the purchase of your system. Our aim is to enable you to experience those special moments that inspire your work.

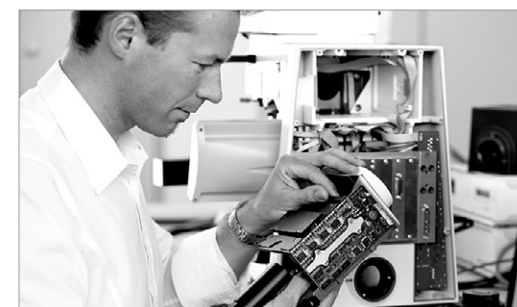
Repair. Maintain. Optimize.

Attain maximum uptime with your microscope. A ZEISS Protect Service Agreement lets you budget for operating costs, all the while reducing costly downtime and achieving the best results through the improved performance of your system. Choose from service agreements designed to give you a range of options and control levels. We'll work with you to select the service program that addresses your system needs and usage requirements, in line with your organization's standard practices.

Our service on-demand also brings you distinct advantages. ZEISS service staff will analyze issues at hand and resolve them – whether using remote maintenance software or working on site.

Enhance Your Microscope System.

Your ZEISS microscope system is designed for a variety of updates: open interfaces allow you to maintain a high technological level at all times. As a result you'll work more efficiently now, while extending the productive lifetime of your microscope as new update possibilities come on stream.



Profit from the optimized performance of your microscope system with services from ZEISS – now and for years to come.

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We make it visible.